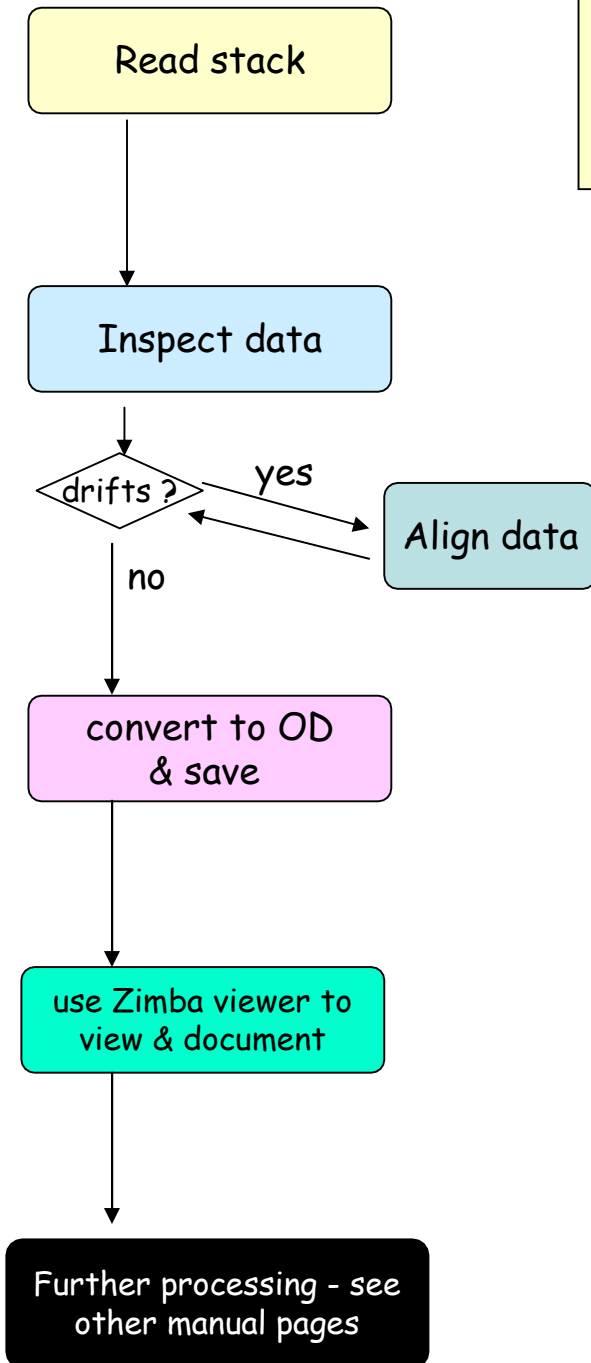


Overview of procedure to process a STXM stack



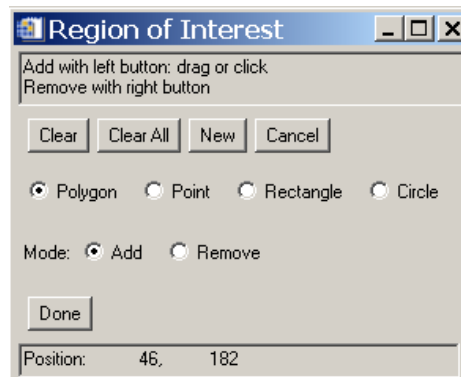
1. `read~STXM(sdf)` or `read~STXM(NeXus)`
2. **browse to file** - click on name
3. **select region** (1 = I, 2 = I_o for example)
4. **select channel** (counter 0, SiPD etc)
5. click 'OK' → reads images
6. **define file name** (e.g. use the index 11_100912034 → 34)
7. **choose zoom** (stack_analyze widget size) choose default

1. **play movie** to check quality - remove bad images etc
2. identify I_o regions (see below)
3. **Dismiss** stack process

1. **read in stack** to `stacks~analyze~Zimba`
OR `stacks~analyze~Jacobsen stack analyze`
2. click on **Auto alignment** (zimba) or
`file~align~start aligning`
3. **save *.aln file** or **save new alignment** (multiple)
4. **read in** to `stack_analyze~axis` binary
5. **select *.aln file**
6. **write out & trim** (*a.ncb) [(*.a2.ncb) if twice etc]

2. **select I_o region** (if one in stack) OR
read I_o from file (recorded separately, or ROI, etc)
- 3 **convert to OD**
4. **SAVE stack** (→ ##jod.ncb) - use systematic naming

1. **read in aligned, OD stack** to `stacks~analyze~Zimba`
2. **skip alignment** - go to **Zstack spectra**
3. **add I region(s)** in areas with different morphologies
4. **screen capture** images, spectra plots to ppt to document
5. **save *.xas** to use as reference spectra in fitting



- Generating quantitative reference spectra
- Fitting stacks to make chemical maps
- Checking quality of fits in spectral space